

A Reassessment of the Lewy Body

Wing Lok Au^{1,2} and Donald B. Calne¹

Abstract- Lewy body has been linked to Parkinson's disease for almost a century, but its significance in neurodegenerative diseases is not known. Whether it is toxic, protective, or just a bystander has been a subject of debate. Recent advances in molecular and genetic works suggest Lewy bodies are not essential for the diagnosis and pathogenesis of Parkinson's disease. Furthermore, the discovery of gene mutations in PARK8, an autosomal-dominant late-onset parkinsonism with pleomorphic pathology, suggests the clinical expression of neurodegenerative diseases depends more on the anatomical pathways affected rather than any particular "pathological marker".

Key Words: Lewy bodies, α -Synuclein, Tau, PARK8, LRRK2

Acta Neurol Taiwan 2018;27:82-89

INTRODUCTION

The presence of Lewy bodies in the substantia nigra has long been considered the pathological hallmark of Parkinson's disease⁽¹⁾. These eosinophilic spherical inclusion bodies were first described by Friederich Heinrich Lewy in 1912⁽²⁾, and were later named after him by Tretiakoff⁽³⁾, who documented the presence of Lewy bodies in the substantia nigra⁽⁴⁾. With better staining techniques, we now appreciate the diffuse nature of these inclusion bodies, not only in terms of topographical distribution within Parkinson's disease⁽⁵⁾, but also their presence in diverse conditions such as subacute sclerosing panencephalitis⁽⁶⁾, Down's syndrome⁽⁷⁾, Hallervorden-Spatz disease⁽⁸⁾, multiple system atrophy⁽⁹⁾, dementia with Lewy bodies⁽⁹⁾, Lewy body variant of

Alzheimer's disease⁽¹⁰⁾, and progressive supranuclear palsy⁽¹¹⁾. Lewy bodies have also been found in the substantia nigra of elderly individuals without neurological disease^(12,13). On the other hand, not all patients with Parkinson's disease have Lewy bodies. They are typically absent in autosomal recessive juvenile-onset Parkinson's disease with parkin gene mutations⁽¹⁴⁻¹⁷⁾. How Lewy bodies are formed, and their possible role in the pathogenesis of Parkinson's disease, remain unclear.

What are Lewy bodies?

The classical description of a Lewy body is an intraneuronal, eosinophilic spherical body with a central core and a pale-staining peripheral halo⁽¹⁸⁾. It measures 8 to 30 μ m in diameter, and stains pink on regular haematoxylin and eosin preparations. Lewy bodies are found

From the ¹Pacific Parkinson's Research Centre, University of British Columbia, Vancouver, Canada; ²Department of Neurology, National Neuroscience Institute, Singapore.

Received November 26, 2017.

Revised and Accepted December 7, 2018.

Reprint requests and corresponding to: Donald B. Calne, MD. Pacific Parkinson's Research Centre, Vancouver Hospital and Health Sciences Centre, Purdy Pavilion, 2221 Wesbrook Mall, Vancouver, B.C. V6T 2B5, Canada.

E-mail: dbcalne@interchange.ubc.ca

in abundance in the surviving neurons of the substantia nigra in Parkinson's disease. They may also be found in the locus ceruleus, dorsal motor nucleus of vagus, nucleus basalis of Meynert, limbic and cortical structures⁽⁵⁾. The cortical Lewy bodies tend to have a diffuse structure without a distinct core and halo⁽¹⁸⁾. Electron-microscopic examination shows the central core to contain granular material, whereas the peripheral halo consists mainly of filamentous structures, radiating from the centre like the spokes of a wheel⁽¹⁹⁾.

Lewy bodies contain a mixture of lipids, proteins, and neurofilaments⁽²⁰⁾. The main constituents are α -synuclein and ubiquitin^(21,22). Lewy bodies without α -synuclein have also been described⁽²³⁾. In the classical well-formed Lewy body, ubiquitin tends to concentrate within the central core, whereas α -synuclein is located mainly in the periphery⁽²⁰⁾. Such a separation of ubiquitin and α -synuclein is not present in the diffuse-type of Lewy body.

Besides α -synuclein and ubiquitin, Lewy bodies also contain ubiquitin-mediated enzymes^(24,25), ubiquitinated proteins⁽²⁴⁾, proteasomes⁽²⁶⁾, proteasome activators, heat-shock proteins, and other centrosome components such as γ -tubulin and pericentrin⁽²⁴⁾. They do not contain synaptophysin, β -synuclein, or γ -synuclein. They have very little or no 20S proteasome α subunits and PA28 activator⁽²⁷⁾.

What is the function of Lewy bodies?

The suspicion that Lewy bodies might be toxic increased when α -synuclein was found in Lewy bodies⁽²¹⁾. This was despite the fact that previous studies did not show an effect of Lewy bodies on cell structure and function^(28,29). Furthermore, Parkinson's disease may occur in the absence of Lewy bodies⁽¹⁵⁾, as demonstrated in post-mortem studies of autosomal recessive young-onset Parkinson's disease with parkin gene mutations^(16,17,30,31).

With improvements in immunohistochemical staining techniques, and advances in transgenic and cell culture studies, interest is now focused on the contents of Lewy bodies, in particular α -synuclein and ubiquitin-proteasome complexes. α -Synuclein is a presynaptic

protein whose physiological function in humans is not known. α -Synuclein knockout mice develop normally with no evidence of Lewy bodies or dopaminergic cell loss⁽³²⁾. However, they had reduced striatal dopamine levels without impairment of locomotive function. They also had abnormal stimulus-dependent release of dopamine, suggesting that under normal circumstances, α -synuclein might play a role as a negative regulator of dopamine release. Other studies showed resistance of MPTP-induced neuronal degeneration in α -synuclein knockout mice^(33,34), suggesting that MPTP-induced neuronal toxicity is dependent on α -synuclein. Whether the mutant or wild-type α -synuclein is neurotoxic is not known. Transgenic mice with both mutant and wild-type α -synuclein develop inclusion bodies in the substantia nigra and extrastriatal neurons⁽³⁵⁻³⁸⁾. In particular, the wild-type α -synuclein transgenic mice may develop locomotive impairment⁽³⁵⁾. Taken together, these findings suggest that α -synuclein associated toxicity is related to a gain of function rather than a loss of function⁽³⁹⁾. The functional gain may be due to an interaction between genetic and environmental factors.

The α -synuclein monomer is normally unfolded, but it can aggregate with other α -synuclein monomers or with other proteins (such as neurofilament proteins) to form Lewy neurites in axons⁽⁴⁰⁾ and Lewy bodies in neurons. Cell culture studies have shown that α -synuclein can undergo polymerization to form fibrils and intermediate oligomeric protofibrils⁽⁴¹⁻⁴³⁾. One current hypothesis is that protofibrils, and not fibrils, are toxic to the cells⁽⁴⁴⁾.

The ubiquitin-proteasome system is important for the processing and degradation of unwanted and possibly cytotoxic proteins⁽⁴⁵⁾. The first step is the activation of ubiquitin by ubiquitin-activating enzyme E1. The high-energy intermediate is then attached covalently to target proteins by other enzymes such as the ubiquitin-conjugating enzyme E2 and the ubiquitin ligase E3 to form a polyubiquitinated protein. The ubiquitinated proteins are then degraded by the 26S proteasome complex, with the release of free and reusable ubiquitin by ubiquitin-recycling enzymes. Lewy bodies contain ubiquitin, ubiquitinated proteins, proteasomes, and other compo-

nents of the ubiquitin-proteasome system⁽²⁴⁻²⁶⁾. This suggests that the formation of Lewy bodies is a cytoprotective mechanism, formed in an attempt to degrade unwanted proteins. *In vitro*, α -synuclein is degraded by the ubiquitin-proteasome pathway⁽⁴⁶⁾. However, recent evidence suggests the ubiquitination of α -synuclein in Lewy bodies might be a pathological event, since unmodified α -synuclein can be degraded by proteasome independent of ubiquitin⁽⁴⁷⁾. The controversy as to whether Lewy bodies are toxic, protective, or just an epiphenomenon, remain unresolved.

The contents of Lewy bodies are similar to those of aggresomes, a microtubule organizing centre that is integral to the regulation of abnormal proteins⁽⁴⁸⁾. Lewy bodies, however, have reduced 20S proteasome α subunits, PA28 activator, and other proteolytic enzymes⁽²⁷⁾. These may limit the normal functioning of the ubiquitin-proteasome system⁽⁴⁹⁾. Perhaps Lewy bodies are dysfunctional aggresomes⁽⁵⁰⁾, formed as a mechanism to contain the unwanted and potentially toxic proteins, unable to be engaged by the degradation process.

Are Lewy bodies essential for the diagnosis of Parkinson's disease?

Parkinson's disease is a heterogeneous group of disorders with multiple causes, both genetic⁽⁵¹⁾ and environmental⁽⁵²⁾. The pathogenic mechanisms may saturate or overwhelm the ubiquitin-proteasome system for handling the unwanted protein load, leading to the formation of Lewy bodies. Toxic intermediate oligomeric protofibrils may form, contributing to the toxicity. Lewy bodies are not necessary for cell death in Parkinson's disease, as in the case of parkin mutation⁽⁵³⁾. Mutation in parkin results in loss of ubiquitin ligase E3 function⁽⁵⁴⁾, which may be necessary for the formation of Lewy bodies. Postmortem studies of patients with parkin mutations generally do not have Lewy bodies^(16,17,30,31) - though there has been one exception⁽⁵⁵⁾. This patient had a compound heterozygous mutation. It is possible that mis-sense mutations of the parkin gene may have some ubiquitin ligase activity, and hence allow formation of Lewy bodies⁽⁵³⁾. In another report, a patient with homozygous exon deletion in the parkin gene did not have Lewy bodies,

but basophilic α -synuclein inclusion bodies were seen in the neuropils of pedunculopontine nucleus⁽³¹⁾. Parkin knockout mice have high dopamine concentrations in the limbic areas, and altered dopamine oxidative metabolism⁽⁵⁶⁾. This evidence indicates that Lewy bodies are not essential for the pathogenesis and diagnosis of Parkinson's disease.

Furthermore, Lewy bodies have been described in a number of other conditions, such as subacute sclerosing panencephalitis⁽⁶⁾, Down's syndrome⁽⁷⁾, Hallervorden-Spatz disease⁽⁸⁾, multiple system atrophy⁽⁹⁾, dementia with Lewy bodies⁽⁹⁾, Lewy body variant of Alzheimer's disease⁽¹⁰⁾, and progressive supranuclear palsy⁽¹¹⁾. They can even be an incidental finding in normal aging^(12,13).

Lewy bodies in Parkinson's disease are not restricted to the substantia nigra, but may be seen outside the nigrostriatal system such as in the locus ceruleus, raphe nucleus, nucleus basalis of Meynert, limbic structures, and neocortex. The pathology of Parkinson's disease seems to begin in the medulla, and ascends in the brainstem to involve the substantia nigra, and eventually the neocortex⁽⁵⁾.

Cortical Lewy bodies

The presence of cortical Lewy bodies has been associated with dementia⁽⁵⁷⁾. So called "dementia with Lewy bodies" is the second most common dementia, characterized by the clinical triad of fluctuating cognition, visual hallucination, and parkinsonism⁽⁵⁸⁾. These symptoms are similar to Parkinson's disease with dementia^(59,60). Clinically, it may be difficult to differentiate between the two conditions, if not for the arbitrary "one-year rule": patients with dementia onset within the first year of diagnosis of parkinsonism are diagnosed to have dementia with Lewy bodies, and those with dementia occurring much later are more likely to have Parkinson's disease with dementia⁽⁵⁸⁾. This distinction seems arbitrary. Pathologically, the separation is less distinct. Both conditions have Lewy bodies in the brainstem, limbic, and cortical structures⁽⁶¹⁻⁶⁴⁾. Subtle differences may exist. For example, Harding et al.⁽⁶⁵⁾ reported more Lewy bodies in the inferotemporal cortex of patients with dementia with Lewy bodies. In Parkinson's disease with dementia, the

Lewy body load is higher in the frontal cortex. Alzheimer's pathology such as neurofibrillary tangles and senile plaques are scattered over the hippocampus, entorhinal cortex, and neocortex in both cases^(61,66). In view of the similarity, it has been suggested that Parkinson's disease with and without dementia, and dementia with Lewy bodies, may represent a spectrum of the same pathological condition^(63,64). It is interesting to note that back in 1923, Lewy had already described the diffuse and cortical distribution of Lewy bodies, and had never suggested making them the pathological hallmark of Parkinson's disease⁽⁴⁾.

Is dementia in Parkinson's disease related to Lewy bodies or Alzheimer's pathology? The presence of Lewy bodies in the cerebral cortex has been linked to the development of dementia^(57,67,68). Some authors reported a correlation between cognitive deficits in Parkinson's disease and the Lewy body load in areas such as the entorhinal cortex, anterior cingulate cortex and frontal cortex^(69,70). Churchyard and Lees⁽⁷¹⁾ reported a significant correlation only with the density of Lewy neurites in the hippocampus. Others, however, did not find any association between clinical symptoms and Lewy body densities⁽⁶¹⁾. Lewy bodies may also be present in the limbic structures and neocortex of Parkinson's disease without dementia⁽⁶⁵⁾. The "Alzheimer's pathology" in Parkinson's disease and dementia with Lewy bodies is usually milder compared to Alzheimer's disease. Alzheimer's pathology up to Braak stage III may be clinically asymptomatic. However, it has been suggested that the additional load of neurofibrillary tangles and senile plaques on a brain laden with Lewy bodies in the limbic system may be additive⁽⁷²⁾.

The presence of tau and α -synuclein together

Tau is a microtubule-associated protein that is present in neuronal and glial inclusions such as neurofibrillary tangles, Pick's bodies, astrocytic plaques, and coiled bodies⁽⁷³⁾. It is found in neurodegenerative diseases such as Alzheimer's disease, Pick's disease, frontotemporal dementia with parkinsonism, progressive supranuclear palsy, and corticobasal ganglia degeneration⁽⁷⁴⁾. These conditions are often grouped together as taupathies.

Likewise, Parkinson's disease, dementia with Lewy bodies, and multiple system atrophy are often grouped together as synucleinopathies⁽⁹⁾. This division, again, is arbitrary. Both tau and α -synuclein have been reported in Alzheimer's disease, Parkinson's disease, dementia with Lewy bodies, and progressive supranuclear palsy^(11,75). The two proteins may co-locate within the same neuron in these conditions^(11,75). Epitope-mapping studies have shown similar α -synuclein staining patterns in Lewy bodies of different neurodegenerative diseases⁽⁷⁶⁾. Using double-immunofluorescence staining techniques, Duda et al.⁽⁷⁷⁾ were able to show the co-existence of tau and α -synuclein within the same inclusion body in familial Parkinson's disease with α -synuclein gene mutation. Similarly, Ishizawa et al.⁽⁷⁸⁾ have shown tau in Lewy bodies of sporadic Parkinson's disease, especially in the locus ceruleus and nucleus basalis of Meynert. Therefore "pathological markers" are not unique to any particular disease, and may co-exist in different neurodegenerative conditions.

Pleomorphism with one etiology

The clinical expressions and pathological changes of neurodegenerative diseases may vary among individuals, even though they may share a common etiology⁽⁷⁹⁾. In the two large families with autosomal-dominant late-onset parkinsonism linked to PARK8 loci on chromosome 12⁽⁸⁰⁻⁸²⁾, the affected members all showed classical parkinsonian features with good response to levodopa therapy. In addition, other features were documented such as dementia, amyotrophy, and supranuclear gaze palsy. Postmortem studies were available in six cases, all without clinical features of dementia. Of these, only one showed classical Lewy bodies in the brainstem. One had diffuse Lewy body disease, another had tau inclusions similar but not diagnostic of progressive supranuclear palsy. The remaining three cases had non-specific nigral cell loss with ubiquitin-positive neuronal inclusions. One of them had mild to moderate "Alzheimer's pathology", the other had mild motor neuron disease. The gene mutations in PARK8 have recently been discovered, and were found to encode for a large, multifunctional protein known as LRRK2 (leucin-rich repeat kinase 2)⁽⁷⁹⁾.

LRRK2 belongs to the ROCO protein family, and contains multiple domains that play important roles in protein-protein interactions and regulation of cellular processes⁽⁸³⁾. In particular, the kinase activity may be crucial for the phosphorylation of tau and α -synuclein. Therefore a single genetic etiology may give rise to different neurodegenerative conditions such as parkinsonism, dementia, and amyotrophy. Neurodegenerative overlap syndrome has previously been reported⁽⁸⁴⁾. Given time, a “new” neurodegenerative disease may appear. The clinical and pathological expressions may depend on interactions between environmental influences and the common genetic defect⁽⁸⁵⁾.

CONCLUSION

We have presented evidence indicating that Lewy bodies are not the “hallmark” of Parkinson’s disease. Furthermore current observations show that the pathological expression of one neurodegenerative etiology can vary markedly from one individual to another. In an attempt to integrate the implications of this finding, we suggest that neurodegenerative mechanisms may be set in motion by a variety of causes, and ultimately converge on a final process of neuronal death. The clinical expression of neurodegeneration will be expressed by the anatomical pathways primarily affected rather than any particular pathological marker. There is even evidence to suggest that with the passage of time, there is convergence of the anatomical pathways attacked by neurodegenerative disorder. If a patient presenting with Alzheimer’s disease lives long enough, the features of Parkinson’s disease will emerge, and vice versa⁽⁸⁴⁾.

REFERENCES

- Gibb WR, Lees AJ. The significance of the Lewy body in the diagnosis of idiopathic Parkinson’s disease. *Neuropathol Appl Neurobiol* 1989;15:27-44.
- Lewy FH. Paralysis agitans: I. Pathologische anatomie. *Handbuch der Neurologie III*. Berlin: Springer, 1912:920-33.
- Tretiakoff C. Contribution a l’étude de l’anatomie pathologique du locus niger et soemmering avec quelques déductions relatives á la pathogénie des troubles du tonus musculaire et de la maladie de Parkinson. University of Paris, 1919.
- Holdorff B. Friedrich Heinrich Lewy (1885-1950) and his work. *J Hist Neurosci* 2002;11:19-28.
- Braak H, Del Tredici K, Rub U, et al. Staging of brain pathology related to sporadic Parkinson’s disease. *Neurobiol Aging* 2003;24:197-211.
- Gibb WR, Scaravilli F, Michund J. Lewy bodies and subacute sclerosing panencephalitis. *J Neurol Neurosurg Psychiatry* 1990;53:710-1.
- Raghavan R, Khin-Nu C, Brown A, et al. Detection of Lewy bodies in Trisomy-21 (Down’s Syndrome). *Can J Neurol Sci* 1993;20:48-51.
- Arawaka S, Saito Y, Murayama S, et al. Lewy body in neurodegeneration with brain iron accumulation type 1 is immunoreactive for alpha-synuclein. *Neurology* 1998;51:887-9.
- Spillantini MG. Parkinson’s disease, dementia with Lewy bodies and multiple system atrophy are alpha-synucleinopathies. *Parkinsonism Relat Disord* 1999;5:157-62.
- Giasson BI, Duda JE, Murray IV, et al. Oxidative damage linked to neurodegeneration by selective alpha-synuclein nitration in synucleinopathy lesions. *Science* 2000;290:985-9.
- Mori H, Oda M, Komori T, et al. Lewy bodies in progressive supranuclear palsy. *Acta Neuropathol (Berl)* 2002;104:273-8.
- Jellinger K. The pathology of parkinsonism. In: Marsden CD, Fahn S, eds. *Movement Disorders 2*. London: Butterworths, 1987:124-65.
- Forno LS, Langston JW. Lewy bodies and aging: relation to Alzheimer’s and Parkinson’s diseases. *Neurodegeneration* 1993;2:19-24.
- Rajput AH, Uitti RJ, Sudhakar S, et al. Parkinsonism and neurofibrillary tangle pathology in pigmented nuclei. *Ann Neurol* 1989;25:602-6.
- Mizuno Y, Hattori N, Matsumine H. Neurochemical and neurogenetic correlates of Parkinson’s disease. *J Neurochem* 1998;71:893-902.
- Mori H, Kondo T, Yokochi M, et al. Pathologic and biochemical studies of juvenile parkinsonism linked to chro-

- mosome 6q. *Neurology* 1998;51:890-2.
17. Hayashi S, Wakabayashi K, Ishikawa A, et al. An autopsy case of autosomal-recessive juvenile parkinsonism with a homozygous exon 4 deletion in the parkin gene. *Mov Disord* 2000;15:884-8.
 18. Lowe J. Lewy bodies. In: Calne DB, ed. *Neurodegenerative Diseases*. Philadelphia: W.B. Saunders, 1994:51-69.
 19. Forno LS. Neuropathology of Parkinson's disease. *J Neuropathol Exp Neurol* 1996;55:259-72.
 20. Gai WP, Yuan HX, Li XQ, et al. In situ and in vitro study of colocalization and segregation of alpha-synuclein, ubiquitin, and lipids in Lewy bodies. *Exp Neurol* 2000;166:324-33.
 21. Spillantini MG, Schmidt ML, Lee VM, et al. Alpha-synuclein in Lewy bodies. *Nature* 1997;388:839-40.
 22. Lennox G, Lowe J, Morrell K, et al. Anti-ubiquitin immunocytochemistry is more sensitive than conventional techniques in the detection of diffuse Lewy body disease. *J Neurol Neurosurg Psychiatry* 1989;52:67-71.
 23. van Duinen SG, Lammers GJ, Maat-Schieman ML, et al. Numerous and widespread alpha-synuclein-negative Lewy bodies in an asymptomatic patient. *Acta Neuropathol (Berl)* 1999;97:533-9.
 24. McNaught KS, Shashidharan P, Perl DP, et al. Aggresome-related biogenesis of Lewy bodies. *Eur J Neurosci* 2002;16:2136-48.
 25. Lowe J, McDermott H, Landon M, et al. Ubiquitin carboxyl-terminal hydrolase (Pgp 9.5) is selectively present in ubiquitinated inclusion-bodies characteristic of human neurodegenerative diseases. *J Pathol* 1990;161:153-60.
 26. Ii K, Ito H, Tanaka K, et al. Immunocytochemical co-localization of the proteasome in ubiquitinated structures in neurodegenerative diseases and the elderly. *J Neuropathol Exp Neurol* 1997;56:125-31.
 27. McNaught KS, Belizaire R, Isacson O, et al. Altered proteasomal function in sporadic Parkinson's disease. *Exp Neurol* 2003;179:38-46.
 28. Gertz HJ, Siegers A, Kuchinke J. Stability of cellsize and nucleolar size in Lewy body containing neurons of substantia-nigra in Parkinson's disease. *Brain Res* 1994;637:339-41.
 29. Tompkins MM, Hill WD. Contribution of somal Lewy bodies to neuronal death. *Brain Res* 1997;775:24-9.
 30. van de Warrenburg BP, Lammens M, Lucking CB, et al. Clinical and pathologic abnormalities in a family with parkinsonism and parkin gene mutations. *Neurology* 2001;56:555-7.
 31. Sasaki S, Shirata A, Yamane K, et al. Parkin-positive autosomal recessive juvenile parkinsonism with alpha-synuclein-positive inclusions. *Neurology* 2004;63:678-82.
 32. Abeliovich A, Schmitz Y, Farinas I, et al. Mice lacking alpha-synuclein display functional deficits in the nigrostriatal dopamine system. *Neuron* 2000;25:239-52.
 33. Dauer W, Kholodilov N, Vila M, et al. Resistance of alpha-synuclein null mice to the parkinsonian neurotoxin MPTP. *Proc Natl Acad Sci USA* 2002;99:14524-9.
 34. Drolet RE, Behrouz B, Lookingland KJ, et al. Mice lacking alpha-synuclein have an attenuated loss of striatal dopamine following prolonged chronic MPTP administration. *Neurotoxicology* 2004;25:761-9.
 35. Masliah E, Rockenstein E, Veinbergs I, et al. Dopaminergic loss and inclusion body formation in alpha-synuclein mice: implications for neurodegenerative disorders. *Science* 2000;287:1265-9.
 36. van der Putten H, Wiederhold KH, Probst A, et al. Neuropathology in mice expressing human alpha-synuclein. *J Neurosci* 2000;20:6021-9.
 37. Giasson BI, Duda JE, Quinn SM, et al. Neuronal alpha-synucleinopathy with severe movement disorder in mice expressing A53T human alpha-synuclein. *Neuron* 2002;34:521-33.
 38. Lee MK, Stirling W, Xu Y, et al. Human alpha-synuclein-harboring familial Parkinson's disease-linked Ala-53 -> Thr mutation causes neurodegenerative disease with alpha-synuclein aggregation in transgenic mice. *Proc Natl Acad Sci USA* 2002;99:8968-73.
 39. Goldberg MS, Lansbury PT Jr. Is there a cause-and-effect relationship between alpha-synuclein fibrillization and Parkinson's disease? *Nat Cell Biol* 2000;2:E115-9.
 40. Braak H, Sandmann-Keil D, Gai W, et al. Extensive axonal Lewy neurites in Parkinson's disease: a novel pathological feature revealed by alpha-synuclein immunocytochemistry. *Neurosci Lett* 1999;265:67-9.
 41. Conway KA, Harper JD, Lansbury PT Jr. Fibrils formed in vitro from alpha-synuclein and two mutant forms linked to Parkinson's disease are typical amyloid. *Biochemistry*

- 2000;39:2552-63.
42. Lee HJ, Lee SJ. Characterization of cytoplasmic alpha-synuclein aggregates. Fibril formation is tightly linked to the inclusion-forming process in cells. *J Biol Chem* 2002; 277:48976-83.
 43. Volles MJ, Lansbury PT Jr. Vesicle permeabilization by protofibrillar alpha-synuclein is sensitive to Parkinson's disease-linked mutations and occurs by a pore-like mechanism. *Biochemistry* 2002;41:4595-602.
 44. Volles MJ, Lansbury PT Jr. Zeroing in on the pathogenic form of alpha-synuclein and its mechanism of neurotoxicity in Parkinson's disease. *Biochemistry* 2003;42:7871-8.
 45. Ciechanover A, Brundin P. The ubiquitin proteasome system in neurodegenerative diseases: sometimes the chicken, sometimes the egg. *Neuron* 2003;40:427-46.
 46. Bennett MC, Bishop JF, Leng Y, et al. Degradation of alpha-synuclein by proteasome. *J Biol Chem* 1999;274: 33855-8.
 47. Tofaris GK, Razaq A, Ghetti B, et al. Ubiquitination of alpha-synuclein in Lewy bodies is a pathological event not associated with impairment of proteasome function. *J Biol Chem* 2003;278:44405-11.
 48. Kopito RR. Aggresomes, inclusion bodies and protein aggregation. *Trends Cell Biol* 2000;10:524-30.
 49. McNaught KS, Olanow CW, Halliwell B, et al. Failure of the ubiquitin-proteasome system in Parkinson's disease. *Nat Rev Neurosci* 2001;2:589-94.
 50. Olanow CW, Perl DP, DeMartino GN, et al. Lewy-body formation is an aggresome-related process: a hypothesis. *Lancet Neurol* 2004;3:496-503.
 51. Le W, Appel SH. Mutant genes responsible for Parkinson's disease. *Curr Opin Pharmacol* 2004;4:79-84.
 52. de la Fuente-Fernandez R, Calne DB. Evidence for environmental causation of Parkinson's disease. *Parkinsonism Relat Disord* 2002;8:235-41.
 53. Hattori N, Mizuno Y. Pathogenetic mechanisms of parkin in Parkinson's disease. *Lancet* 2004;364:722-4.
 54. Shimura H, Hattori N, Kubo S, et al. Familial Parkinson disease gene product, parkin, is a ubiquitin-protein ligase. *Nat Genet* 2000;25:302-5.
 55. Farrer M, Chan P, Chen R, et al. Lewy bodies and parkinsonism in families with parkin mutations. *Ann Neurol* 2001;50:293-300.
 56. Itier JM, Ibanez P, Mena MA, et al. Parkin gene inactivation alters behaviour and dopamine neurotransmission in the mouse. *Hum Mol Genet* 2003;12:2277-91.
 57. Kosaka K, Yoshimura M, Ikeda K, et al. Diffuse type of Lewy body disease: progressive dementia with abundant cortical Lewy bodies and senile changes of varying degree--a new disease? *Clin Neuropathol* 1984;3:185-92.
 58. McKeith IG, Galasko D, Kosaka K, et al. Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB): report of the consortium on DLB international workshop. *Neurology* 1996;47:1113-24.
 59. Aarsland D, Ballard C, Larsen JP, et al. A comparative study of psychiatric symptoms in dementia with Lewy bodies and Parkinson's disease with and without dementia. *Int J Geriatr Psychiatry* 2001;16:528-36.
 60. Ballard CG, Aarsland D, McKeith I, et al. Fluctuations in attention: PD dementia vs DLB with parkinsonism. *Neurology* 2002;59:1714-20.
 61. Gomez-Tortosa E, Newell K, Irizarry MC, et al. Clinical and quantitative pathologic correlates of dementia with Lewy bodies. *Neurology* 1999;53:1284-91.
 62. Harding AJ, Halliday GM. Cortical Lewy body pathology in the diagnosis of dementia. *Acta Neuropathol (Berl)* 2001;102:355-63.
 63. Colosimo C, Hughes AJ, Kilford L, et al. Lewy body cortical involvement may not always predict dementia in Parkinson's disease. *J Neurol Neurosurg Psychiatry* 2003; 74:852-6.
 64. Hishikawa N, Hashizume Y, Yoshida M, et al. Clinical and neuropathological correlates of Lewy body disease. *Acta Neuropathol (Berl)* 2003;105:341-50.
 65. Harding AJ, Broe GA, Halliday GM. Visual hallucinations in Lewy body disease relate to Lewy bodies in the temporal lobe. *Brain* 2002;125(Pt 2):391-403.
 66. Apaydin H, Ahlskog JE, Parisi JE, et al. Parkinson disease neuropathology: later-developing dementia and loss of the levodopa response. *Arch Neurol* 2002;59:102-12.
 67. Gibb WR, Esiri MM, Lees AJ. Clinical and pathological features of diffuse cortical Lewy body disease (Lewy body dementia). *Brain* 1987;110 (Pt 5):1131-53.
 68. Hurtig HI, Trojanowski JQ, Galvin J, et al. Alpha-synuclein cortical Lewy bodies correlate with dementia in Parkinson's disease. *Neurology* 2000;54:1916-21.

69. Mattila PM, Rinne JO, Helenius H, et al. Alpha-synuclein-immunoreactive cortical Lewy bodies are associated with cognitive impairment in Parkinson's disease. *Acta Neuropathol (Berl)* 2000;100:285-90.
70. Kovari E, Gold G, Herrmann FR, et al. Lewy body densities in the entorhinal and anterior cingulate cortex predict cognitive deficits in Parkinson's disease. *Acta Neuropathol (Berl)* 2003;106:83-8.
71. Churchyard A, Lees AJ. The relationship between dementia and direct involvement of the hippocampus and amygdala in Parkinson's disease. *Neurology* 1997;49:1570-6.
72. Braak H, Braak E, Yilmazer D, et al. Neurofibrillary tangles and neuropil threads as a cause of dementia in Parkinson's disease. *J Neural Transm Suppl* 1997;51:49-55.
73. Dickson DW. Tau and synuclein and their role in neuropathology. *Brain Pathol* 1999;9:657-61.
74. Buee L, Delacourte A. Comparative biochemistry of tau in progressive supranuclear palsy, corticobasal degeneration, FTDP-17 and Pick's disease. *Brain Pathol* 1999;9:681-93.
75. Schmidt ML, Martin JA, Lee VM, et al. Convergence of Lewy bodies and neurofibrillary tangles in amygdala neurons of Alzheimer's disease and Lewy body disorders. *Acta Neuropathol (Berl)* 1996;91:475-81.
76. Lippa CF, Schmidt ML, Lee VM, et al. Alpha-synuclein in familial Alzheimer disease: epitope mapping parallels dementia with Lewy bodies and Parkinson disease. *Arch Neurol* 2001;58:1817-20.
77. Duda JE, Giasson BI, Mahon ME, et al. Concurrence of alpha-synuclein and tau brain pathology in the Contursi kindred. *Acta Neuropathol (Berl)* 2002;104:7-11.
78. Ishizawa T, Mattila P, Davies P, et al. Colocalization of tau and alpha-synuclein epitopes in Lewy bodies. *J Neuropathol Exp Neurol* 2003;62:389-97.
79. Zimprich A, Biskup S, Leitner P, et al. Mutations in LRRK2 cause autosomal-dominant parkinsonism with pleomorphic pathology. *Neuron* 2004;44:601-7.
80. Wszolek ZK, Vieregge P, Uitti RJ, et al. German-Canadian family (family A) with parkinsonism, amyotrophy, and dementia -- Longitudinal observations. *Parkinsonism Relat Disord* 1997;3:125-39.
81. Wszolek ZK, Pfeiffer RF, Tsuboi Y, et al. Autosomal dominant parkinsonism associated with variable synuclein and tau pathology. *Neurology* 2004;62:1619-22.
82. Zimprich A, Muller-Myhsok B, Farrer M, et al. The PARK8 locus in autosomal dominant parkinsonism: confirmation of linkage and further delineation of the disease-containing interval. *Am J Hum Genet* 2004;74:11-9.
83. Shen J. Protein kinases linked to the pathogenesis of Parkinson's disease. *Neuron* 2004;44:575-7.
84. Uitti RJ, Berry K, Yasuhara O, et al. Neurodegenerative 'overlap' syndrome: clinical and pathological features of Parkinson's disease, motor neuron disease, and Alzheimer's disease. *Parkinsonism Relat Disord* 1995;1:21-34.
85. Uitti RJ, Calne DB, Dickson DW, et al. Is the neuropathological 'gold standard' diagnosis dead? Implications of clinicopathological findings in an autosomal dominant neurodegenerative disorder. *Parkinsonism Relat Disord* 2004;10:461-3.